

**Supporting Information for**

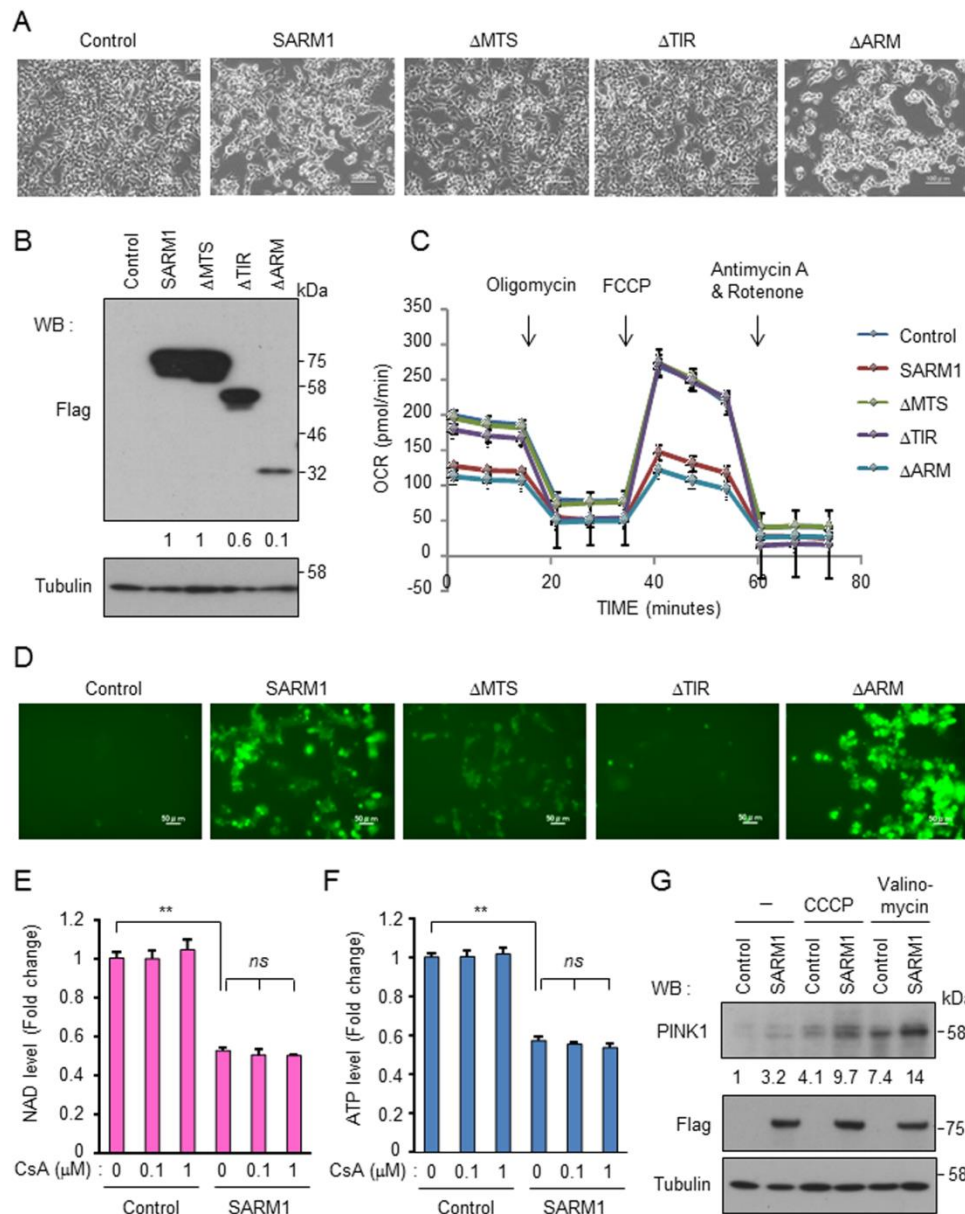
**JNK-mediated phosphorylation of SARM1 regulates NAD<sup>+</sup> cleavage activity to inhibit mitochondrial respiration**

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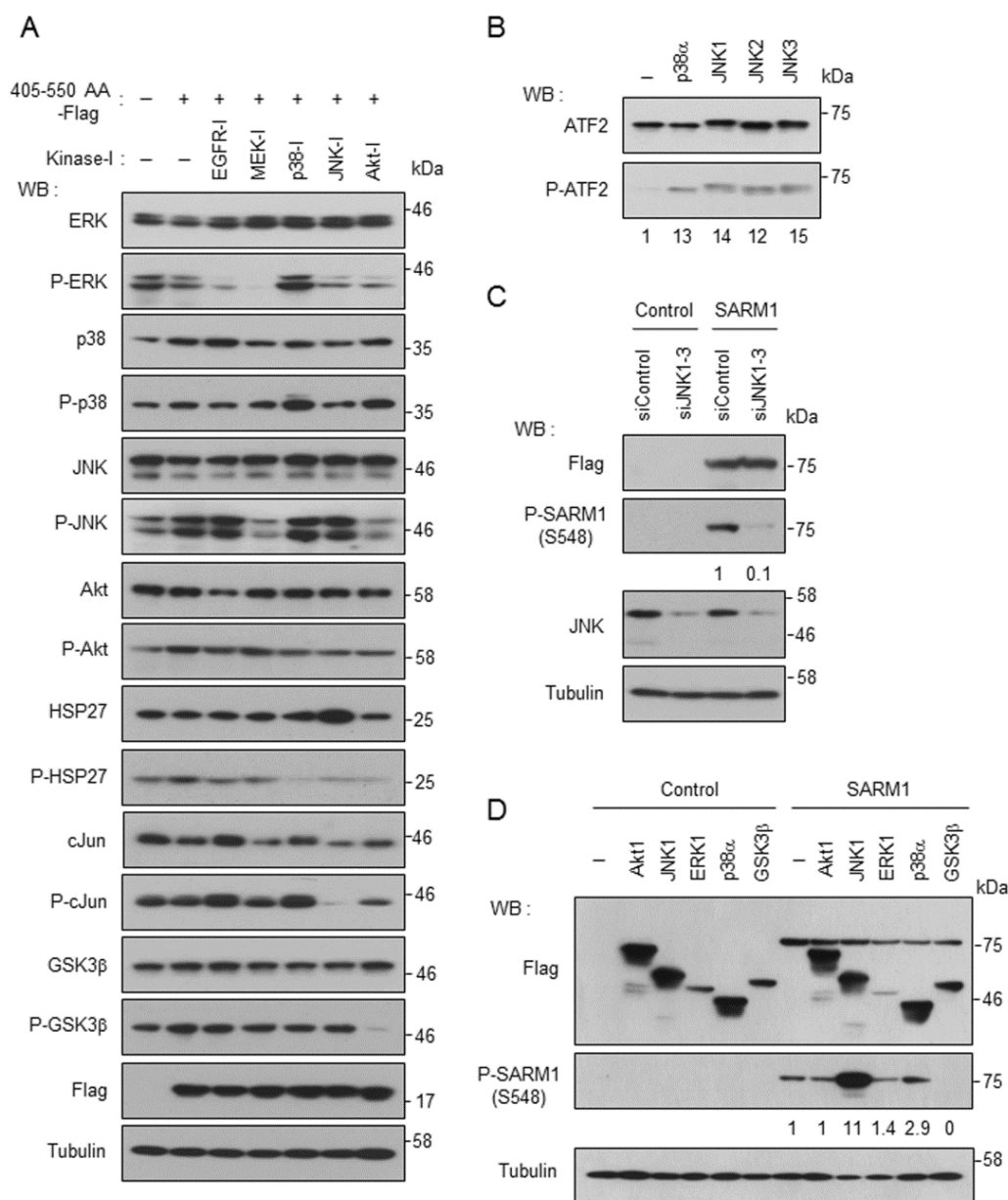
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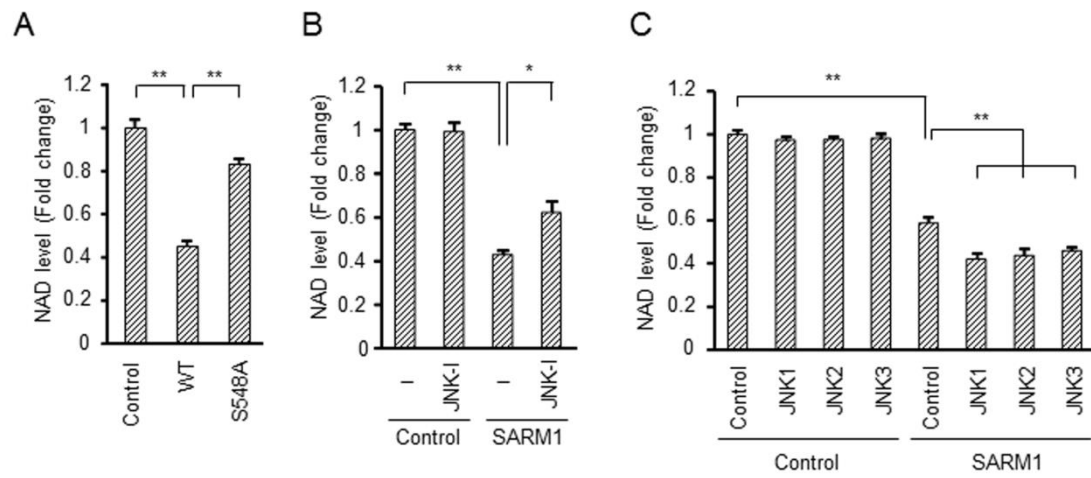
Figs. S1 to S5



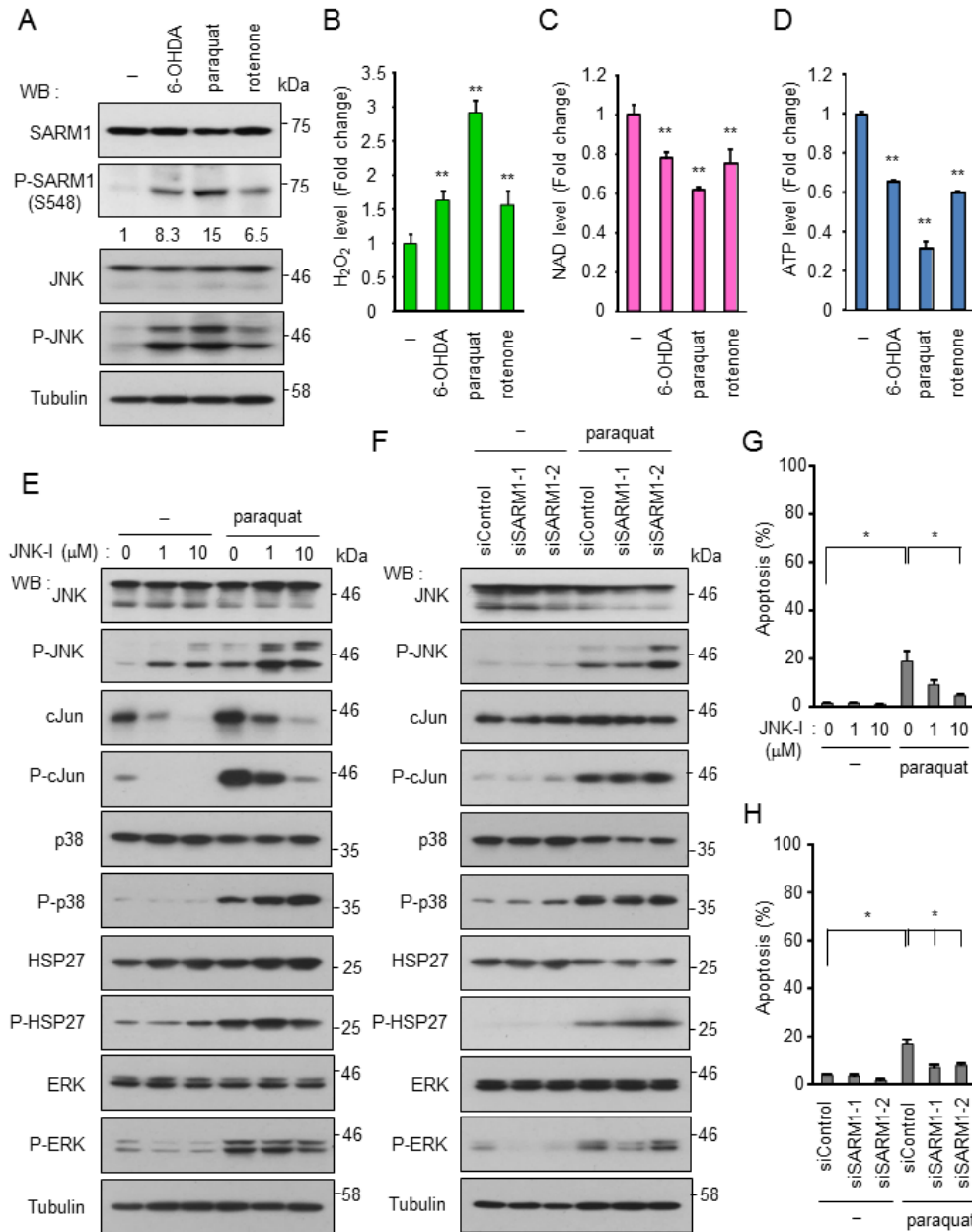
**Figure S1. SARM1 overexpression hampers mitochondrial function.** (A) Phase contrast micrographs of HEK293T cells expressing SARM1 constructs. Bar, 100  $\mu$ m. (B) Expression level of SARM1 constructs in HEK293T cells. Flag-tagged SARM1 constructs were densitometrically quantified (Flag/ Tubulin). (C) To analyze OCR, 1.5  $\mu$ M oligomycin, 0.25  $\mu$ M FCCP and 0.5  $\mu$ M rotenone/antimycin A were used. (D) ROS production was analyzed by CM-H<sub>2</sub>DCFDA staining. Bar, 50  $\mu$ m. (E and F) Cyclosporin A (CsA), a PTP inhibitor, did not affect SARM1-induced NAD<sup>+</sup> and ATP reduction. (G) SARM1 overexpression enhances PINK1 accumulation on depolarized mitochondria. HEK293T cells were transfected with pDNA. At 24 h post-transfection, the cells were treated with 10  $\mu$ M CCCP or 10  $\mu$ M valinomycin and further incubated for 3 h. PINK1 were quantified by densitometry (PINK1/ Tubulin).



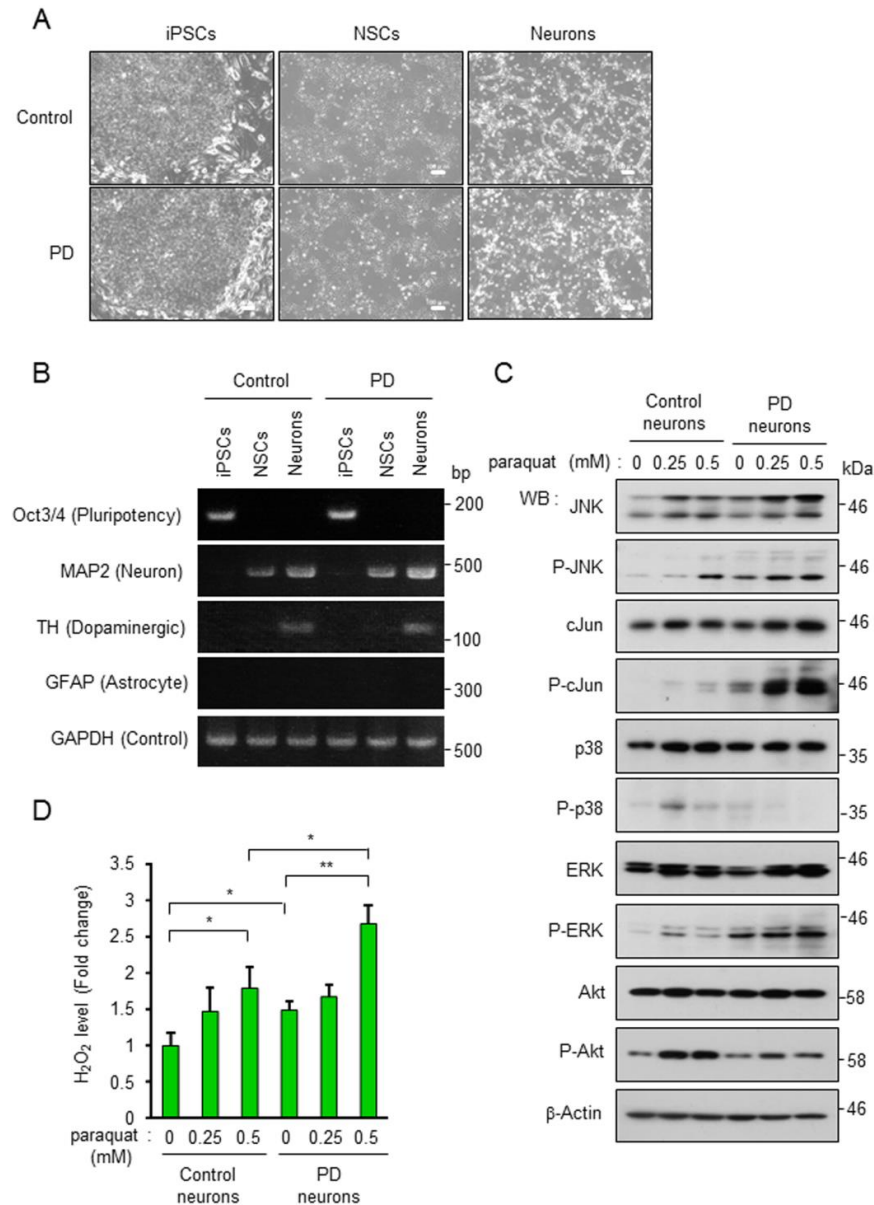
**Figure S2. JNK regulates SARM1 phosphorylation.** (A) Confirmation of the inhibitory effect of kinase inhibitors. The experiment was performed under same conditions to those in Fig. 3A. (B) Confirmation of the catalytic activity of kinases. (C) Down-regulation of SARM1 phosphorylation by knock-down of JNK. Control siRNA or JNK siRNAs (mixtures of siRNA targeting JNK1, 2 and 3) was transfected in HEK293T cells. At 24 h post-transfection, the cells were transfected with control or SARM1-Flag pDNA and further incubated for 24 h. (D) Up-regulation of SARM1 phosphorylation by overexpression of JNK. The protein expression was confirmed by anti-Flag antibody. Phosphorylated proteins were quantified by densitometry (Phosphorylated protein/ total protein).



**Figure S3. JNK-mediated phosphorylation of SARM1 affects intracellular NAD<sup>+</sup> level.**  
 (A-C) The indicated constructs were transfected in HEK293T cells for 24 h. the intracellular NAD<sup>+</sup> levels were measured by the NAD/NADH-Glo assay.



**Figure S4. Induction of SARM1 phosphorylation by oxidative stresses.** (A) Detection of endogenous SARM1 phosphorylation. SH-SY5Y cells were treated with 20  $\mu$ M 6-OHDA, 1 mM paraquat or 1  $\mu$ M rotenone for 24 h. (B-D) Oxidative stresses change levels of ROS, NAD<sup>+</sup> and ATP. (E) Phosphorylation of MAPK cascade proteins under paraquat and JNK-I treatment conditions. The experiment was performed under same conditions to those in Fig. 5A. (F) Phosphorylation of MAPK cascade proteins under paraquat treatment and SARM1 down-regulation conditions. The experiment was performed under same conditions to those in Fig. 5D. (G and H) Apoptotic cells were identified after staining with Hoechst33342. \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ .



**Figure S5. Induction of NSCs and matured neurons from human control iPS cells and PD patient-derived iPS cells.** (A) Representative images of control and PD-derived iPSCs, NSCs and neurons. Bar, 100  $\mu$ m. (B) Marker analysis of iPSCs, NSCs and neurons. RT-PCR analysis showed almost the same expression levels of pluripotency markers Oct3/4 in iPSC, neuronal marker MAP2 in NSC and neurons, and dopaminergic neuron marker TH in neurons between control and PD-derived cells. The astrocyte marker GFAP showed no expression and GAPDH was used as a control. (C) Phosphorylation of MAPK cascade proteins of neurons under paraquat treatment conditions. (D) H<sub>2</sub>O<sub>2</sub> levels of Control neurons and PD neurons under paraquat treatment conditions. \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ .